

EPA 533 Sampling Instructions

Prior to sampling:

- Have ice available to fill the bag labelled "ICE Bag" once the samples are collected.
- Be prepared to mail the sample (on ice) directly after collecting it or drop it off at the lab.
- Review the Best Practices-PFAS Sample Collection brochure provided with your kit.

Sampling Source/Location:

- Sample 1 and 2 must be collected from the same water source, at the same time.
- If the sampling source is filtered, consider removing the filter or sampling before the filter.

Sampling Method:

- If sampling water from a faucet, spigot, or wellhead, run the system for approximately 5 minutes before collecting the sample.
- If sampling a body of water, try to collect the visually "cleanest" sample possible. Avoid sampling particulates and murky water.
- A "Field Blank" (FRB) must be utilized during collection. The Field Blank is comprised of clean PFAS free water provided by the analytical laboratory. It must be opened and placed near your sampling location prior to collecting your first sample and must be closed after collecting your second sample. The Field Blank must be put on ice and shipped to the analytical laboratory with your samples. The purpose of the Field Blank is to detect any potential PFAS source in the air or dust (not from the water sample) at the sampling location.

Collecting the sample:

1. Open the cooler by removing the lid and take out the contents.
2. Fill out the Chain of Custody Form. We need this form to process your sample. Don't forget to add all collection times.
3. Wash your hands, dry them, and put on the provided nitrile gloves prior to sampling.
4. Turn on the water source and run the system for 5 minutes to flush the system.
5. Open the FRB bag, remove the FRB sample, remove the cap and set the FRB sample near the water source you are testing. Be sure to place it in a location where it will not accidentally tip over. Avoid touching the inside of the sample bottle or cap.
6. Open the sample bag and remove sample bottles 1 and 2 (replicate).
7. Open sample bottle 1. Avoid touching the inside of the sample bottle or cap.
8. Do not rinse the sample bottle, it has a preservative (ammonium acetate) in it that is required to be a specific concentration.
9. Fill the bottle with exactly 250 mL of cold water from the water source. Do not over fill or under fill, 250 mL volume is marked on the bottle.
10. After filling, seal the bottle tightly, turn the opening away from your face and shake it until the preservative is dissolved, usually for a minute.
11. Repeat steps 7-10 with sample 2 (replicate). Do not turn off the water in between samples.
12. Replace the cap on the FRB sample and place in the "FRB" bag. Avoid touching the inside of the sample bottle or cap.
13. Then place sample 1 and sample 2 (replicate) back in the "Sample Bag."
14. Remove gloves and discard.
15. Place the sample and FRB bags into the cooler.
16. Fill the provided ice bag with ice and place it in the cooler. Do not place the sample in the ice bag.
17. Put the lid back on the cooler.
18. Place the complete Chain of Custody Form on top of the cooler lid and ensure it is signed. We can't accept unsigned COCs. Be sure to fill in the collection time for every sample, including the Field Blank (FRB).
19. Seal the box with tape.
20. Adhere the provided return label to the box if using.
21. Ship or deliver the collected samples to the analytical lab as soon as possible. Samples must be shipped on ice. Samples are valid if any ice remains in the cooler when it is received at the laboratory or bottles are received within 2 days of collection and below 10 °C.

SPEC LAB will send the test results to the email listed on the Chain of Custody Form.